

## Small Molecule Integrin Antagonists in Cancer Therapy

Mayra Paolillo<sup>1,\*</sup>, Marika A. Russo<sup>1</sup>, Massimo Serra<sup>2</sup>, Lino Colombo<sup>2</sup> and Sergio Schinelli<sup>1</sup>

<sup>1</sup>*Dipartimento di Farmacologia Sperimentale ed Applicata and* <sup>2</sup>*Dipartimento di Chimica Farmaceutica, Università di Pavia – Italy*

**Abstract:** Integrins are a large family of dimeric receptors composed by  $\alpha$  and  $\beta$  subunits that, once bound to extracellular matrix (ECM) proteins, regulate a variety of cellular processes such as cell motility, migration, and proliferation. The integrins transduce signals from inside-out and outside-in the cell, thus representing the cellular link to the external environment. For these properties, integrin activation has been involved in pathological processes like tumor growth and metastasis formation. Recent advances in the elucidation of the crystallographic structures of the  $\alpha v \beta 3$  and  $\alpha II \beta 3$  integrins are promoting studies focused to the search of small molecule antagonists that can block the integrin binding to ECM and inhibit the biological effects exerted by these receptors. In this review we will focus on small molecule antagonists of  $\alpha v \beta 3$  and  $\alpha v \beta 5$  integrin as tools for cancer therapy while other integrins will only be briefly mentioned. Cilengitide (cyclic peptidic  $\alpha v \beta 3$  and  $\alpha v \beta 5$  antagonist) is currently in clinical trials for anti cancer therapy. Combination of integrin  $\alpha v \beta 3$  antagonists and other traditional therapeutic approaches may represent a future strategy to inhibit tumor growth and metastasis spreading.

**Key Words:** Integrin antagonists, RGD-peptides, metastasis, anoikis.

### INTRODUCTION

A fundamental aspect related to cancer therapy is the control of tumor cells proliferative and aggressive behaviour. Different strategies have been tested but the main problem encountered by the most recent therapeutic approaches is represented by tumor heterogeneity. Tumors greatly vary in their cell composition and genotype and even tumors belonging to the same histological classification may bear different mutations. This heterogeneity makes the identification of new pathways and promising therapeutic targets difficult and slow. At present, the cellular growth control is generally achieved by traditional chemotherapeutic agents but new strategies supporting the classical antiproliferative approach, like antiangiogenic agents, are explored [1]. In the last decade, studies concerning the interactions between tumor cells and the extracellular environment highlighted the possibility of identifying molecules that may represent interesting therapeutic targets for tumor cell proliferation and motility control [2]. Neoplastic growth is a complex process resulting from interactions among different cell types present in the tumor mass such as tumor cells, cells present in the tumoral niche, fibroblast and endothelial cells, that contribute to the composition of the extracellular matrix (ECM). ECM is a complex and dynamic structure that differs in protein composition and regulates cellular processes like cell migration, proliferation and differentiation [2].

ECM components can be broadly classified in glycosylated proteins and proteoglycans. The ECM proteins are represented by the fibronectin molecule and by two families, laminins and collagens, that include several distinct struc-

tural and functional members [3]. The ECM proteins exert their effect by binding cell surface receptors, ubiquitously expressed in almost any mammalian cell, denominated integrins.

The integrin receptor family have been identified as key regulators of different cellular processes related to tumoral growth like cell adhesion and migration, cell proliferation and neo-angiogenesis. Several integrin antagonists are in clinical trial for cancer therapy and in this review we will examine some features of the most promising small molecule integrin antagonists together with their potential therapeutic use.

### THE INTEGRIN RECEPTOR FAMILY

Integrins are heterodimeric glycoprotein receptors formed by the combination of different 18  $\alpha$ -subunits and 8  $\beta$ -subunits that give rise to 24 distinct receptors [4]. Integrin expression, though ubiquitous in human tissues, varies with cell types;  $\alpha$  and  $\beta$  subunits are expressed in different combinations and exhibit ligand specificities [4]. The integrins expressed by cells, therefore, control the ECM components with which the cell can interact. Several types of integrin dimers have been identified in cancer cells [5] where they appear to play key roles mainly in proliferative and metastatic processes. The  $\alpha 5 \beta 1$ ,  $\alpha v \beta 3$  and  $\alpha v \beta 5$  are widely expressed in several cancer types and recognize the tripeptide sequence Arg-Gly-Asp (RGD), found in many ECM proteins [6]. The structure of  $\alpha v \beta 3$ , determined in unliganded [7] and ligand-bound states [8], showed a crevice formed by the ectodomain of the  $\beta$  chain [9], that is the RGD-like peptide binding site. Interestingly, integrin expression changes during the transition from a non-neoplastic to a neoplastic state suggesting that alterations of the adhesion properties in cancer cells may be involved in the early steps of metastasis formation [10,11].

\*Address correspondence to this author at the Dipartimento di Farmacologia Sperimentale ed Applicata, Università di Pavia, Italy, E-mail: mayra.paolillo@unipv.it

## INTEGRIN MEDIATED SIGNALLING

Through the extracellular domain, integrins bind ECM proteins while inside the cell the short cytoplasmic tail binds to the cytoskeleton [9,12]. This is a bi-directional interaction because the cytoskeleton proteins modulate the affinity of the integrin-ECM binding and, conversely, perturbations of integrin binding to ECM change the shape and protein assembly of the cytoskeleton. This tight linkage between cells and ECM is achieved through the interaction of the short cytoplasmic domain with proteins like talin1, tensin, vinculin,  $\alpha$ -actinin and filamin [13]. The adapter protein paxillin may link talin to the integrin  $\alpha$ -tails contributing to the formation of adhesion complexes and enhancing resistance of ECM-integrin-talin to mechanical stresses [13]. Focal adhesions are integrin-mediated adhesion complexes that may vary in composition according to integrins and matrix components. The mechanism underlying cell motility has been clarified in detail, although not completely on the molecular structure level: actin filaments apply force on the ECM-integrin complex leading to the transformation of focal complexes into focal adhesions indicating the direction of cell movement. Afterwards, the integrins located on the trailing edge of the cell are internalized to be recycled to the leading edge where new interactions with ECM are formed [13]. This mechanism is required to cross tissue layers while other mechanisms regulate cell migration in the interstitial space [14].

Integrin binding to ECM ligands transduce signals to the tyrosin kinase Src that is constitutively bound to the integrin  $\beta$  cytoplasmic tail. In clustered integrins, Src transphosphorylation leads to recruitment of tyrosine phosphatases [15,16] and to activation of kinases like phosphatidylinositol-4-phosphate 5 kinase type  $\gamma$  (PIPKI $\gamma$ ) or FAK [17]. These signals contribute to the activation and recruitment of focal adhesion proteins like vinculin and talin and, through phosphorylation of intracellular transducers like p130Cas or caveolin, lead to the activation of the classical ERK pathways implicated in survival and proliferation signalling [18]. It should be considered that growth factors receptors (GFRs) are influenced by the adhesion state of the cells and FAK-stimulated signalling cascade has been shown to modulate the ability of GFRs to stimulate ERK phosphorylation and cell cycle phases transition [19]. This combination of adhesion and survival signals triggered by integrin-ECM binding is considered to be the mechanism by which integrin-mediated adhesion promotes cell survival.

When wrong interactions between cells and ECM proteins cause the cell detachment from ECM, a kind of apoptosis called "anoikis" may be induced. Anoikis, homelessness in greek, indicates a condition in which proliferation and survival signals are suppressed and apoptotic caspase-dependent and independent processes start [20]. It was clearly demonstrated that integrins suppress anoikis by FAK activation in a variety of cell types and that integrin attachment to fibronectin counteracts the effect of mitochondrial pro-apoptotic proteins [21]; on the other hand, cilengitide and RGD peptides induce anoikis in human brain cancer and endothelial cells [22-24], suggesting that blocking of  $\alpha$ v $\beta$ 3 and  $\alpha$ v $\beta$ 5 receptors may represent a promising strategy to suppress apoptotic resistance in cancer cells.

## INTEGRINS AND CANCER

Classical chemotherapeutic anticancer therapies try to hit the tumoral mass or to inhibit the growth of satellite foci but the main point is that 90% of death of cancers patients is caused by metastasis and not by the solid tumors [25]. While the tumoral growth has been widely studied and the steps that lead cells towards cancerogenic transformation process have been deeply investigated, the metastasis formation has received less attention and the molecular events regulating this complex process are only partially known. The metastatic process requires several complex steps that can be summarized in loss of cellular adhesion, increased cell motility and invasiveness, entry and survival in the circulation, crossing new tissue barriers, and colonization of distant sites [26,27].  $\alpha$ v $\beta$ 3 integrins have been implicated in metastasis formation, specifically during adhesion of circulating tumor cells to the vascular endothelium and crossing the endothelial cell layer [28,29].  $\alpha$ v $\beta$ 3 and  $\alpha$ v $\beta$ 5 appear to be directly involved in metastasis formation in different cancer types like colon cancer, metastatic melanoma and breast cancer [2]. In glioma samples,  $\alpha$ v $\beta$ 3 expression has been detected in glial tumor cells and correlates with tumor grade, suggesting that these receptors may be important players in the highly infiltrative behaviour of high grade gliomas [30].

## INTEGRIN ANTAGONISTS

Because of their role in metastatic and infiltrative processes,  $\alpha$ v $\beta$ 3 and  $\alpha$ v $\beta$ 5 represent interesting targets for new anticancer strategies. In addition, integrins expressed by endothelial cells participate in angiogenesis. Indeed, integrin  $\alpha$ v $\beta$ 3 expression was found to be increased in proliferating vascular endothelial cells and for this reason it was thought to be involved in pathological angiogenesis [31]. These findings suggested the potential use of RGD-like  $\alpha$ v $\beta$ 3 inhibitors as antiangiogenic agents in cancer therapy. However, results from *in vivo* studies are controversial because while the antiangiogenic effect was beneath expectations, RGD-like inhibitors appear to affect glioma cell survival directly [23]. Therefore, the effect of integrin antagonists on tumor progression is still difficult to define and phase I and phase II studies often propose different mechanisms for the observed effects.

A different family of integrin inhibitors is represented by antibodies that bind to  $\alpha$ v $\beta$ 3. In particular, two antibodies have been successfully developed and are currently in clinical trial: Vitaxin (MEDI-523) is in phase I for the treatment of stage IV metastatic melanoma and androgen-independent prostate cancer and Abegrin (MEDI-522), a modification of MEDI-523, is in phase I for patients with solid tumors (abegrin, vitaxin). However, since the discussion of these inhibitors is beyond the scope of this review, for a more detailed discussion we suggest to see Tucker, 2006 [32].

## PEPTIDES AND PEPTIDE-MIMETIC "SMALL MOLECULES" INTEGRIN INHIBITORS.

These compounds are peptide-mimetics carrying in their structure the RGD sequence [33,34] or nonpeptidic derivatives capable to interact with the integrin receptors through functional groups mimicking the guanidine and carboxylic moieties present in the side chain of arginine and aspartic acid, respectively.

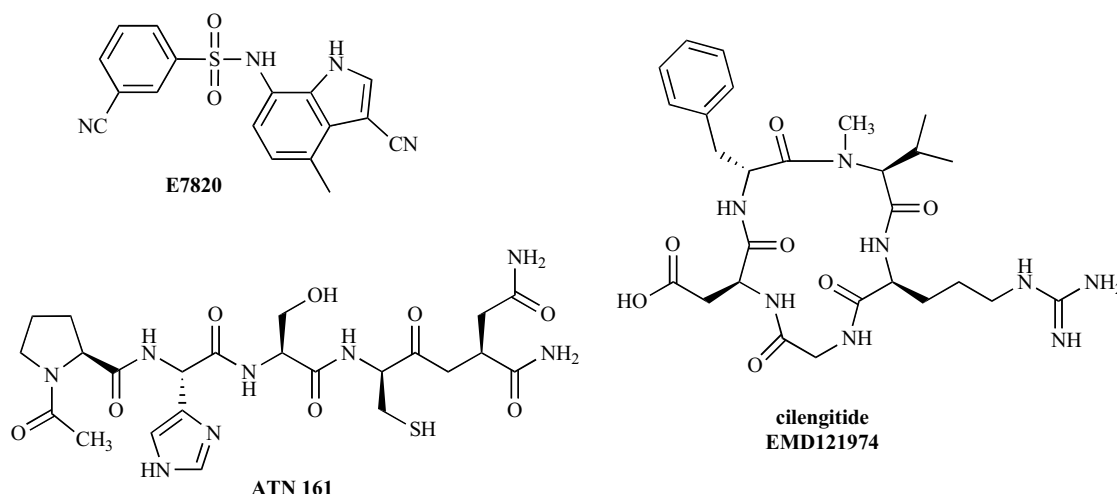


Fig. (1).

**CILENGITIDE (EMD 121974)**

The first integrin antagonist to be tested in humans has been Cilengitide (EMD 121974), a cyclic peptide belonging to the RGD-peptide family that binds to the described crevice of the  $\beta$  chain thus blocking the binding to ECM [35]. Cyclic RGD pentapeptides with constrained backbone conformations have first been developed as highly active and selective ligands for the  $\alpha v\beta 3$  integrin receptor [36-38]. Extensive ligand-based drug design studies led to the highly active  $\alpha v\beta 3$  selective first generation inhibitor *cyclo*(Arg-Gly-Asp-D-Phe-Val) [36,39-41]. The systematic derivatization of the lead peptide resulted in the *N*-methylated cyclopeptide EMD 121974, *cyclo*(Arg-Gly-Asp-D-Phe-NMeVal), as one of the most active compounds. The X-ray analysis of the complex  $\alpha v\beta 3$ -EMD121974 shows that the ligand interacts mainly through electrostatic interactions [8,42]. Arg and Asp of the RGD form a “charged clamp” that binds regions with opposite charges in the protein: Asp interacts with a metal cation in the MIDAS region of the  $\beta$  subunit and Arg with two residues of aspartic acid (Asp218 and Asp150) located in the  $\alpha$  subunit.

In preclinical studies, cilengitide has been shown to block glioblastoma (GBM) growth in nude mice [43] while other studies report that cilengitide in combination with radiotherapy decreased the tumor proliferation rate in ovarian cancer

xenografts [44]. In this case, the decreased proliferation rate was not attributed to anti-angiogenic activity but to a proapoptotic effect. Cilengitide, in combination with temozolomide, was also reported to exert antiproliferative effects on melanoma cells *in vitro* and a reduction of melanoma growth, compared to the methylating agent alone, was also observed *in vivo* [45].

The efficacy of RGD peptides in clinical trials has been demonstrated (Table 1), though in some cases results are controversial. First data demonstrated good tolerability and low toxicity in patients, which prompted the initiation of further studies using a combination with chemo- or radiotherapy [45,46]. However, a recent study in patients with recurrent GBM has shown that cilengitide monotherapy, though well tolerated, has modest antitumor activity [47]. Another study reports that co-application of cilengitide and gemcitabina resulted in partial tumor remission in a patient with highly vascularized head and neck cancer and in this case the anticancer activity was attributed to an anti-angiogenic effect [48]. In patients with different solid tumors Cilengitide monotherapy gave no partial or complete responses [49], phase II studies have been designed to evaluate Cilengitide treatments in patients with non metastatic androgen-independent prostate cancer (NCC 6735) and patients with metastatic prostate cancer (NCC 6372). Recently, RGD cyclic peptides have been used as carriers for cytotoxic drugs

**Table 1. Clinical Trials of Small Molecule  $\alpha v\beta 3$  and  $\alpha v\beta 5$  Inhibitors**

Name		Highest Phase Reached	Ref.
Cilengitide (EMD 121974)	glioblastoma	Phase III	[80]
ATN-161	malignant glioma	Phase I	[81]
	renal cell cancer	Phase II	[82]
E7820	solid tumor, lymphoma	Phase I	[83]
	colorectal cancer	Phase II	[84]

[50,51], to specifically target cancer cells and tumor blood vessels. Furthermore, suitably radiolabeled RGD peptides have been developed for non invasive imaging of integrin expression during tumor angiogenesis, growth, and spread in humans [52,53].

### ATN-161

ATN-161 is an N-acetylated, amidated, non RGD-based, linear pentapeptide (Ac-PHSCN-NH<sub>2</sub>) in phase II clinical trial in patients with solid tumors (see Table 1). It is a non-competitive inhibitor of the fibronectin "synergy region" PHSRN sequence. This fibronectin region increases the affinity and specificity of the RGD-mediated binding thus making integrin-fibronectin interaction stronger. In ATN-161 a cysteine residue has been introduced for arginine along with peptide acetylation and amidation, in order to yield a product with acceptable pharmaceutical properties.

An unregulated invasive response to the PHSRN sequence may contribute significantly to the growth, survival and metastasis of established tumors [54]. Recent studies show that ATN-161 binds to integrin beta subunits of  $\alpha 5\beta 1$  and  $\alpha v\beta 3$  subtypes and displays antitumorogenic and antimetastatic activities in different cancer types [55,56].

### E7820

This molecule is an aromatic sulfonamide derivative (*N*-(3-Cyano-4-methyl-1H-indol-7-yl)-3-cyanobenzene-sulfonamide). E7820 cannot be considered a classical integrin antagonist since it does not bind to  $\alpha$  or  $\beta$  integrin subunits, nevertheless an inhibitory effect on  $\alpha 2$  integrin mRNA expression in endothelial cells was reported [57]. E7820 has demonstrated anti-angiogenic activity in both in-vitro and in-vivo angiogenesis models and this, in turn, leads to a decrease in cancer cell proliferation rate in different cancer models [57,58]. The antiangiogenic effects have been ascribed to inhibition of vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF) activity. Inhibition of  $\alpha 2$  expression might be the mechanism underlying this effect. The drug is currently in phase II clinical development (see Table 1).

## NEW MOLECULES

### Peptides

During the last decade other peptidomimetic compounds have been synthesized and tested for their ability in blocking

RGD-mediated cell adhesion. Several turn mimetics [59,60] and sugar amino acids were incorporated into the lead structure cyclo(RGDf[NMe]V) by replacing the D-Phe-[NMe]Val dipeptide unit. A common feature of these derivatives was the presence of a 15 membered macrocycle, identical to that of EMD121974. Studies on peptide secondary structure lead to the synthesis of several 6,5- and 7,5-fused 2-oxo-1-azabicyclo[X.3.0]alkane amino acids possessing the general structure 1 (Fig. 2) [61]. These scaffolds can be regarded as conformationally restricted substitutes for Ala-Pro and Phe-Pro dipeptide units [50,62]. All the bicyclic compounds include a natural L-Pro residue, but vary in the lactam ring size and in the configuration at the fusion carbon and at the NH<sub>2</sub>-bearing center. Computational and spectroscopic studies have revealed that these scaffolds can mimic reverse-turn motifs as those investigated by Haubner and coworkers [39], although there is a dependence of the turn-inducing ability on the lactam ring size and stereochemistry [63]. Replacement of the D-Phe-[NMe]Val dipeptide in the reference compound cyclo(RGDf[NMe]V) with such azabicycloalkane derivatives, showing different reverse-turn mimetic properties, constrained the RGD sequence into specific conformations and provided the required activity and selectivity for integrin antagonism. For a more complete figure of turn mimetics for the D-Phe-Val units see Weide *et al.* [33].

A library of RGD-containing cyclic pseudopentapeptides was then synthesized, allowing the achievement of some specific high-affinity  $\alpha v\beta 3$  and  $\alpha v\beta 5$  integrin antagonists. Among the peptides tested, ST1646 and compound 2 showed the highest affinity towards  $\alpha v\beta 3$  receptor, inhibiting [<sup>125</sup>I]echistatin binding to  $\alpha v\beta 3$  with an IC<sub>50</sub> value of  $3.7 \pm 0.6$  nM and of  $6.4 \pm 0.1$  nM respectively, that are comparable to EMD121974 IC<sub>50</sub> values ( $18.9 \pm 3.1$  nM) in the same assay.

In preliminary experiments, compound 2 has shown an inhibitory effect on cell attachment and cell migration assays, probably mediated by FAK inhibition, in human glioblastoma cells (Russo *et al.*, manuscript in preparation). These data highlight the possibility of *in vivo* experiments to further clarify the anticancer activity of this compound.

Another series of peptide antagonists was recently patented [64]. These compounds are a hybrid cyclopeptide derivative embodying pyrrolidine or piperidine-based amino acid grafted onto a RGD tripeptide sequence and belongs to

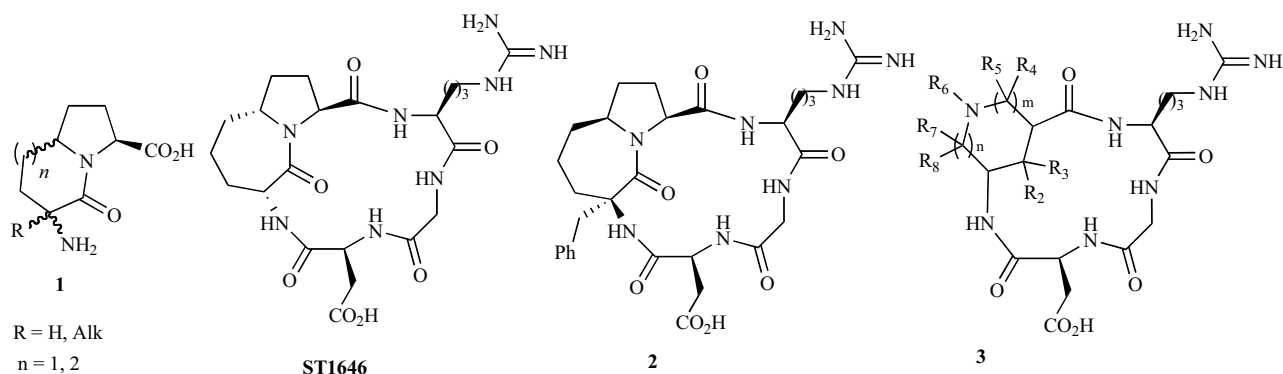


Fig. (2).

a family of compounds of general formula 3. The structural novelty is represented by pseudopeptide 14-atom macrocycles more constrained than the aforementioned cyclopentapeptides and these compounds exhibited low-nanomolar and even picomolar antagonist activity towards the  $\alpha v\beta 3$  and  $\alpha v\beta 5$  integrins.

### Non Peptidic

The need for antagonists with higher bioavailability and lower molecular weight has prompted several research groups to develop small constrained nonpeptidic molecules mimicking the RGD motif. Many of the structures proposed consist of a polyfunctionalized rigid core, linked to appendages corresponding to arginine and aspartic acid side chains. Both the basicity and the length of the arginine-mimic group showed to be essential for binding interaction with the integrin receptor. Moreover, the presence of a carboxylic function mimicking the aspartic acid of RGD is an essential feature to create anionic interaction with the MIDAS cationic region.

Many heterocyclic scaffolds have been employed to hold the acidic and the basic ends of the molecule at the appropriate distance and with the suitable conformation for binding interaction [34]. Here we only report compounds which strongly bind to integrins involved in the tumoral process and which therefore are promising candidates for future applications in cancer therapy.

Recently, the aforementioned azabicyclo[4.3.0]alkanes were functionalized with guanidinium or carboxylate groups on both cyclic components [65]. The most active lactam of the series was **4**, bearing a  $-\text{CH}_2\text{CO}_2\text{H}$  group on the proline ring and an arginine residue linked to the amino group at the 3-position.

In their continuous efforts focused at finding highly active and selective  $\alpha v\beta 3$  integrin antagonists, Kessler and co-workers designed new RGD mimetics in which the glycine was replaced either by an aza-glycine residue (see **5a**, Fig. 3) [66] or an azacarba analogue (compound **5b**) [67]. The aspartic acid was mimicked by various  $\beta$ -amino acids or glutaric acids carrying different substituents at the  $\beta$ -position. A guanidinic function linked to the GD-mimic by either aromatic or aliphatic spacers completed the structure. The most active compound of the series was compound **6**, that showed an excellent  $\text{IC}_{50}$  value in the subnanomolar range (0.1 nM). By carrying out some structural modifications on compounds of general formula 5, the same research group showed that a set of integrin specific inhibitors can be designed around a conserved aza-glycine backbone [68]. Compound **7** was the first specific, low molecular weight  $\alpha v\beta 6$  peptidomimetic inhibitor described in the literature.

The  $\alpha v\beta 6$  integrin receptor is up-regulated in tumor proliferation [69,70] and in a number of pathological processes such as inflammatory events, wound healing, cell migration and proliferation. This class of inhibitors is therefore of great interest for future applications in human pathologies such as cancer and osteoporosis.

Another integrin which has recently been drawn into the focus of research is the  $\alpha 5\beta 1$  subtype, whose pro-angiogenic function has been clearly demonstrated [71]. Recently, the

synthesis of new nonpeptide  $\alpha v\beta 3/\alpha 5\beta 1$  integrin dual antagonists containing 5,6-dihydropyridin-2-one scaffolds was reported. The dihydropyridinone heterocycle was converted into a series of potential integrin antagonists changing the length, the structure and the position of the side chains bearing the acidic and the basic appendages. The biological activity of the various derivatives has been evaluated by testing their ability to inhibit cell attachment mediated by  $\alpha v\beta 3$  and  $\alpha 5\beta 1$  integrins in cell adhesion assays. The enantiomerically pure derivative **8** had excellent affinity towards both receptors [72].

The hint for the design of new  $\alpha 5\beta 1$  ligands came from the homology model of the  $\alpha 5\beta 1$  receptor [73]. On the basis of these studies Heckmann *et al.* [74] synthesized a series of compounds based on a tyrosine scaffold that has already been successfully employed in the integrin field [75]. Compound **9** showed the best  $\alpha 5\beta 1$  affinity (0.7 nM) and good selectivity against  $\alpha v\beta 3$  (300-fold). In another paper [76], the same authors used the structure-activity informations provided by a tyrosine-based compounds library to design new  $\alpha 5\beta 1$  aza-glycine-based ligands with affinities of  $\sim 1$  nM and selectivities that exceed  $10^4$ -fold. The activity on  $\alpha 5\beta 1$  and selectivity against  $\alpha v\beta 3$  seems to be strongly connected to the presence of a *S* configured, alpha-amino acid as carrier of the carboxyl moiety and an aromatic amide with an ortho-dimethyl-substitution pattern (compound **10**).

Stragies *et al.* synthesized a new class of  $\alpha 5\beta 1$  pyrrolidine-based antagonists of general formula **11** [77]. Integrin selectivity was tuned, in agreement with the observations reported by Hackmann *et al.*, by switching from a sulphonamide moiety to a mesitylene amide moiety for R3. Moreover, an increase of activity by a factor of 4-6 was achieved by introducing 4-methoxy-2-aminopyridine as a basic group. Compounds **11a** and **11b** showed subnanomolar  $\alpha 5\beta 1$  activity and very high selectivity toward other related integrin receptors.

Different groups investigated the use of a central heterocycle core in order to maintain the aspartate and the guanidine mimetics in the appropriate orientation to modulate the pharmacokinetic profile of the  $\alpha v\beta 3$  antagonists.

Several indol-1-yl propionic acids, containing a variety of basic moieties at the 5-position as well as substitutions alpha and beta to the carboxy terminus, were reported [78]. The most active compound **12** carries a 3-pyridyl substituent on the carboxylic chain and a tetrahydronaphthyridine moiety at the 5-position of the indole ring. This derivative possesses a number of favorable properties including good oral bioavailability and a subnanomolar activity towards  $\alpha v\beta 3$  receptor.

In other studies the synthesis of benzazepine derivatives is reported (compound **13**). These molecules are highly potent, orally active (in rats)  $\alpha v\beta 3$  antagonists. An extensive study aimed at increasing the lipophilicity of these derivatives revealed that a trifluoro-ethyl-substituent placed on the 2-position considerably improved oral bioavailability in rats. Moreover, incorporation of the 6-(*N*-methylamino)pyridine Arg mimetic had a significant impact on biological activity [79].

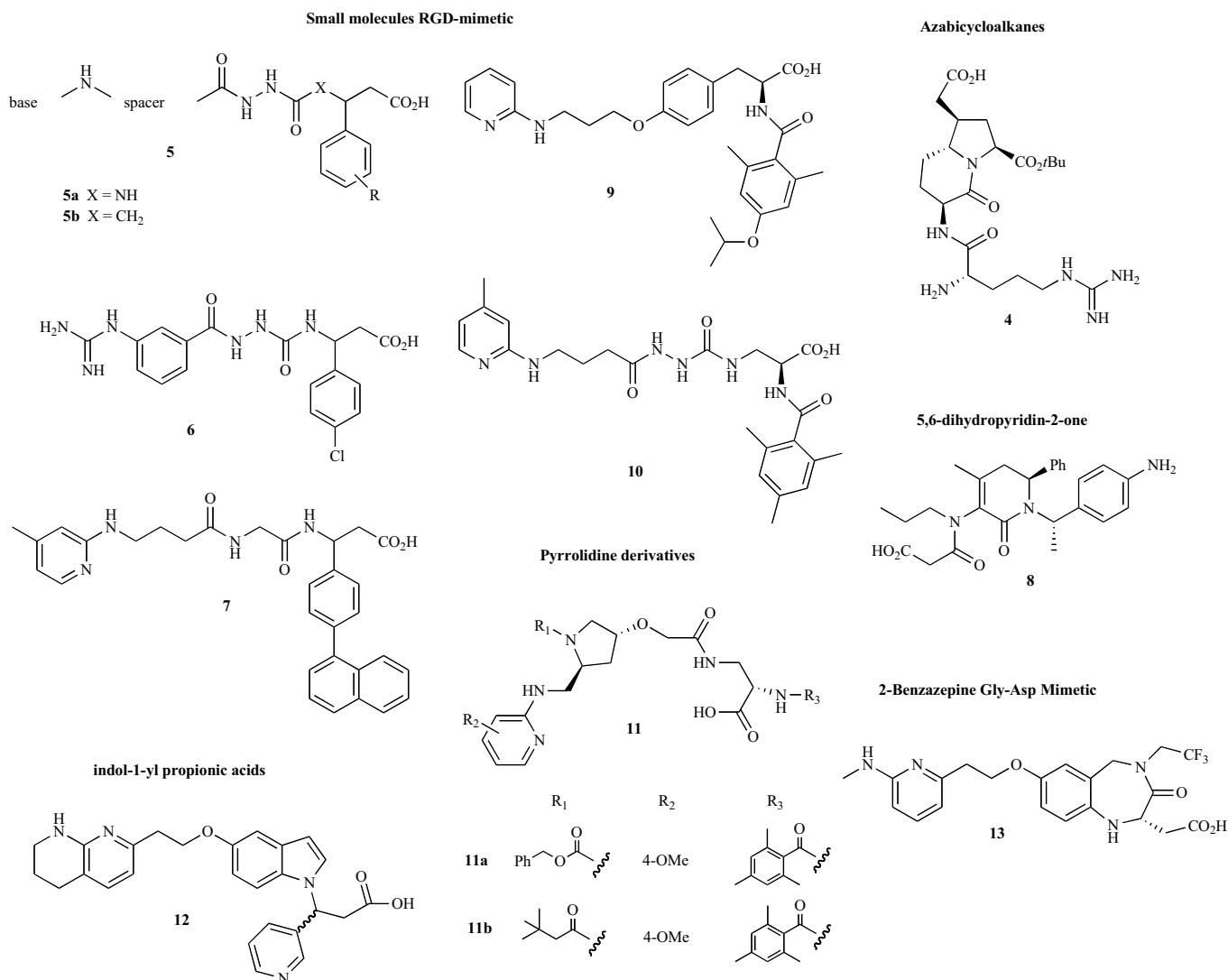


Fig. (3).

## CONCLUSION

The results so far obtained from clinical trials with  $\alpha v \beta 3$  and  $\alpha v \beta 5$  antagonists suggest that these molecules, though exerting both anti-angiogenic and direct anticancer activity, when administered alone, achieve too weak results as anti-cancer agents. On the contrary, very promising results come from trials in which the compounds are associated to chemo- or radio-therapy.

Ongoing studies highlight other interesting applications for  $\alpha v \beta 3$  and  $\alpha v \beta 5$  antagonist, like selective delivery of cytostatic drugs or diagnostic uses in combination with imaging techniques. The development of new small molecule integrin antagonists with good bioavailability could give an important contribution to the most important target of anti-cancer therapy: the arrest of the metastasis spread.

## REFERENCES

- Davis, I.D.; Desai, J. Clinical use of therapies targeting tumor vasculature and stroma. *Curr. Cancer Drug Targets*, **2008**, *8*, 498-508.
- Dittmar, T.; Heyder, C.; Gloria-Maercker, E.; Hatzmann, W.; Zanker, K.S. Adhesion molecules and chemokines: the navigation system for circulating tumor (stem) cells to metastasize in an organ-specific manner. *Clin. Exp. Metastasis*, **2008**, *25*, 11-32.
- Berrier, A.L.; Yamada, K.M. Cell-matrix adhesion. *J. Cell Physiol.*, **2007**, *213*, 565-573.
- Hehlhans, S.; Haase, M.; Cordes, N. Signalling via integrins: implications for cell survival and anticancer strategies. *Biochim. Biophys. Acta*, **2007**, *1775*, 163-180.
- Moschos, S.J.; Drogowski, L.M.; Reppert, S.L.; Kirkwood, J.M. Integrins and Cancer. *Oncology*, **2007**, *21*, 13-20.
- Ruoslahti, E.; Pierschbacher, M.D. Arg-Gly-Asp: a versatile cell recognition signal. *Cell*, **1986**, *44*, 517-518.
- Xiong, J.P.; Stehle, T.; Diefenbach, B.; Zhang, R.; Dunker, R.; Scott, D.L.; Joachimiak, A.; Goodman, S.L.; Arnaout, M.A. Crystal structure of the extracellular segment of integrin alpha V beta 3. *Science*, **2001**, *294*, 339-345.
- Xiong, J.P.; Stehle, T.; Zhang, R.; Joachimiak, A.; Frech, M.; Goodman, S.L.; Arnaout, M.A. Crystal structure of the extracellular segment of integrin alpha V beta 3 in complex with an Arg-Gly-Asp ligand. *Science*, **2002**, *296*, 151-155.
- Arnaout, M.A.; Goodman, S.L.; Xiong, J.P. Structure and mechanics of integrin-based cell adhesion. *Curr. Opin. Cell Biol.*, **2007**, *19*, 495-507.
- Kikkawa, H.; Kaihou, M.; Horaguchi, N.; Uchida, T.; Imafuku, H.; Takiguchi, A.; Yamazaki, Y.; Koike, C.; Kuruto, R.; Kakiuchi, T.; Tsukada, H.; Takada, Y.; Matsuura, N.; Oku, N. Role of integrin alpha(v)beta3 in the early phase of liver metastasis: PET and IVM analyses. *Clin. Exp. Metastasis*, **2002**, *19*, 717-725.

- [11] Enns, A.; Korb, T.; Schlüter, K.; Gassmann, P.; Spiegel, H.U.; Senninger, N.; Mitjans, F.; Haier, J. Alphavbeta5-integrins mediate early steps of metastasis formation. *Eur. J. Cancer*, **2005**, *41*, 1065-72.
- [12] Zhu, J.; Luo, B.H.; Xiao, T.; Zhang, C.; Nishida, N.; Springer, T.A. Structure of a complete integrin ectodomain in a physiologic resting state and activation and deactivation by applied forces. *Mol. Cell*, **2008**, *32*, 849-861.
- [13] Stupack, D.G. The biology of integrins. *Oncology*, **2007**, *21*, 6-12.
- [14] Lämmermann, T.; Bader, B.L.; Monkley, S.J.; Worbs, T.; Wedlich-Saldner, R.; Hirsch, K.; Keller, M.; Förster, R.; Critchley, D.R.; Fässler, R.; Sixt, M. Rapid leukocyte migration by integrin-independent flowing and squeezing. *Nature*, **2008**, *453*, 51-55.
- [15] Baker, M.W.; Peterson, S.M.; Macagno, E.R. The receptor phosphatase HmLAR2 collaborates with focal adhesion proteins in filopodial tips to control growth cone morphology. *Dev. Biol.*, **2008**, *320*, 215-225.
- [16] Carlucci, A.; Gedressi, C.; Lignitto, L.; Nezi, L.; Villa-Moruzzi, E.; Avvedimento, E.V.; Gottesman, M.; Garbi, C.; Feliciello, A. Protein-tyrosine phosphatase PTPD1 regulates focal adhesion kinase autophosphorylation and cell migration. *J. Biol. Chem.*, **2008**, *283*, 10919-29.
- [17] Ling, K.; Doughman, R.L.; Iyer, V.V.; Firestone, A. J.; Bairstow, S.F.; Mosher, D.F.; Schaller, M.D.; Anderson, R. A. Tyrosine phosphorylation of type I{gamma} phosphatidylinositol phosphate kinase by Src regulates an integrin-talin switch. *J. Cell Biol.*, **2003**, *163*, 1339-49.
- [18] Assoian, R.K.; Klein, E.A. Growth control by intracellular tension and extracellular stiffness. *Trends Cell Biol.*, **2008**, *18*, 347-352.
- [19] Bill, H.M.; Knudsen, B.; Moores, S.L.; Muthuswamy, S.K.; Rao, V.R.; Brugge, J.S.; Miranti, C.K. Epidermal growth factor receptor-dependent regulation of integrin-mediated signaling and cell cycle entry in epithelial cells. *Mol. Cell Biol.*, **2004**, *24*, 8586-99.
- [20] Chiarugi, P.; Giannoni, E. Anoikis: a necessary death program for anchorage-dependent cells. *Biochem. Pharmacol.*, **2008**, *76*, 1352-64.
- [21] Gilmore, A.P. Anoikis. *Cell Death Differ.*, **2005**, *12*, 1473-77.
- [22] Maubant, S.; Saint-Dizier, D.; Boutillon, M.; Perron-Sierra, F.; Casara, P.J.; Hickman, J.A.; Tucker, G.C.; Van Obberghen-Schilling, E. Blockade of alpha v beta3 and alpha v beta5 integrins by RGD mimetics induces anoikis and not integrin-mediated death in human endothelial cells. *Blood*, **2006**, *108*, 3035-44.
- [23] Oliveira-Ferrer, L.; Hauschild, J.; Fiedler, W.; Bokemeyer, C.; Nippgen, J.; Celik, I.; Schuch, G. Cilengitide induces cellular detachment and apoptosis in endothelial and glioma cells mediated by inhibition of FAK/src/AKT pathway. *J. Exp. Clin. Cancer Res.*, **2008**, *27*, 86-90.
- [24] Taga, T.; Suzuki, A.; Gonzalez-Gomez, I.; Gilles, F.H.; Stins, M.; Shimada, H.; Barsky, L.; Weinberg, K.L.; Laug, W.E. alpha v-Integrin antagonist EMD 121974 induces apoptosis in brain tumor cells growing on vitronectin and tenascin. *Int. J. Cancer*, **2002**, *98*, 690-697.
- [25] Gupta, G.P.; Massagué, J. Cancer metastasis: building a framework. *Cell*, **2006**, *127*, 679-695.
- [26] Chambers, A.F.; Groom, A.C.; MacDonald, I.C. Dissemination and growth of cancer cells in metastatic sites. *Nat. Rev. Cancer*, **2002**, *2*, 563-572.
- [27] Fidler, I.J. The pathogenesis of cancer metastasis: the 'seed and soil' hypothesis revisited. *Nat. Rev. Cancer*, **2003**, *3*, 453-458.
- [28] Felding-Habermann, B.; O'Toole, T.E.; Smith, J.W.; Fransvea, E.; Ruggeri, Z.M.; Ginsberg, M.H.; Hughes, P.E.; Pampori, N.; Shattil, S.J.; Saven, A.; Mueller, B.M. Integrin activation controls metastasis in human breast cancer. *Proc. Natl. Acad. Sci.*, **2001**, *98*, 1853-58.
- [29] Wang, H.; Fu, W.; Im, J.H.; Zhou, Z.; Santoro, S.A.; Iyer, V.; DiPersio, C.M.; Yu, Q.C.; Quaranta, V.; Al-Mehdi, A.; Muschel, R.J. Tumor cell alpha3beta1 integrin and vascular laminin-5 mediate pulmonary arrest and metastasis. *J. Cell Biol.*, **2004**, *164*, 935-941.
- [30] Schnell, O.; Krebs, B.; Wagner, E.; Romagna, A. Expression of integrin alphavbeta3 in gliomas correlates with tumor grade and is not restricted to tumor vasculature. *Brain Pathol.*, **2008**, *18*, 378-386.
- [31] Hodivala-Dilke, K. alphavbeta3 integrin and angiogenesis: a moody integrin in a changing environment. *Curr. Opin. Cell Biol.*, **2008**, *20*, 514-519.
- [32] Tucker, G.C. Integrins: molecular targets in cancer therapy. *Curr. Oncol. Rep.*, **2006**, *8*, 96-103.
- [33] Weide, T.; Modlinger, A.; Kessler, H. Spatial screening for the identification of the bioactive conformation of integrin ligands. *Top. Curr. Chem.*, **2007**, *272*, 41-50.
- [34] Meyer, A.; Auernheimer, J.; Modlinger, A.; Kessler, H. Targeting RGD Recognizing Integrins: Drug Development, Biomaterial Research, Tumor Imaging and Targeting. *Curr. Pharm. Des.*, **2006**, *12*, 2723-47.
- [35] Dechantsreiter, M. A.; Planker, E.; Matha, B.; Lohof, E.; Hölzemann, G.; Jonczyk, A.; Goodman, S.L.; Kessler, H. N-methylated cyclic RGD peptides as highly active and selective alpha3beta3 integrin antagonists. *J. Med. Chem.*, **1999**, *42*, 3033-40.
- [36] Aumailley, M.; Gurrath, M.; Müller, G.; Calvete, J.; Timpl, R.; Kessler, H. Arg-Gly-Asp constrained within cyclic pentapeptides Strong and selective inhibitors of cell adhesion to vitronectin and laminin fragment P1. *FEBS Lett.*, **1991**, *291*, 50-54.
- [37] Gurrath, M.; Müller, G.; Kessler, H.; Aumailley, M.; Timpl, R. Conformation/activity studies of rationally designed potent anti-adhesive RGD peptides. *Eur. J. Biochem.*, **1992**, *210*, 911-921.
- [38] Müller, G.; Gurrath, M.; Kessler, H.; Timpl, R. Dynamic Forcing, a Method for Evaluating Activity and Selectivity Profiles of RGD (Arg-Gly-Asp) Peptides. *Angew. Chem. Int. Ed.*, **1992**, *31*, 326-328.
- [39] Haubner, R.; Gratias, R.; Diefenbach, B.; Goodman, S.L.; Jonczyk, A.; Kessler, H. Structural and functional aspects of rgd-containing cyclic pentapeptides as highly potent and selective integrin alpha3beta3 antagonists. *J. Am. Chem. Soc.*, **1996**, *118*, 7461-72.
- [40] Wermuth, J.; Goodman, S.L.; Jonczyk, A.; Kessler, H. Stereoisomerism and biological activity of the selective and superactive alpha3beta3 integrin inhibitor cyclo-(RGDFV-) and its retro-inverso peptide. *J. Am. Chem. Soc.*, **1997**, *119*, 1328-35.
- [41] Haubner, R.; Finsinger, D.; Kessler, H. Stereoisomeric Peptide Libraries and Peptidomimetics for Designing Selective Inhibitors of the alpha3beta3 Integrin for a New Cancer Therapy. *Angew. Chem. Int. Ed.*, **1997**, *36*, 1374-89.
- [42] Gottschalk, K.E.; Kessler, H. The Structures of Integrins and Integrin-Ligand Complexes: Implications for Drug Design and Signal Transduction. *Angew. Chem.*, **2002**, *41*, 3767-74.
- [43] Yamada, S.; Bu, X.Y.; Khankaldyyan, V.; Gonzales-Gomez, I.; McComb, J.G.; Laug, W.E. Effect of the angiogenesis inhibitor Cilengitide (EMD 121974) on glioblastoma growth in nude mice. *Neurosurgery*, **2006**, *59*, 1304-12.
- [44] Zhao, H.; Wang, J.C.; Sun, Q.S.; Luo, C.L.; Zhang, Q. RGD-based strategies for improving antitumor activity of paclitaxel-loaded liposomes in nude mice xenografted with human ovarian cancer. *J. Drug Target*, **2008**, *24*, 1.
- [45] Tentori, L.; Dorio, A.S.; Muzi, A.; Lacal, P.M.; Ruffini, F.; Navarra, P.; Graziani, G. The integrin antagonist cilengitide increases the antitumor activity of temozolomide against malignant melanoma. *Oncol. Rep.*, **2008**, *19*, 1039-43.
- [46] Reardon, D.A.; Nabors, L.B.; Stupp, R.; Mikkelsen, T. Cilengitide: an integrin-targeting arginine-glycine-aspartic acid peptide with promising activity for glioblastoma multiforme. *Expert Opin. Investig. Drugs*, **2008**, *17*, 1225-35.
- [47] Reardon, D.A.; Fink, K.L.; Mikkelsen, T.; Cloughesy, T.F.; O'Neill, A.; Plotkin, S.; Glantz, M.; Ravin, P.; Raizer, J.J.; Rich, K.M.; Schiff, D.; Shapiro, W.R.; Burdette-Radoux, S.; Dropcho, E.J.; Wittemer, S.M.; Nippgen, J.; Picard, M.; Nabors, L.B. Randomized phase II study of cilengitide, an integrin-targeting arginine-glycine-aspartic acid peptide, in recurrent glioblastoma multiforme. *J. Clin. Oncol.*, **2008**, *26*, 5610-17.
- [48] Raguse, J.D.; Gath, H.J.; Bier, J.; Riess, H.; Oettle, H. Cilengitide (EMD 121974) arrests the growth of a heavily pretreated highly vascularised head and neck tumour. *Oral Oncol.*, **2004**, *40*, 228-230.
- [49] Hariharan, S.; Gustafson, D.; Holden, S.; McConkey, D.; Davis, D.; Morrow, M.; Basche, M.; Gore, L.; Zang, C.; O'Bryant, C.L.; Baron, A.; Gallemann, D.; Colevas, D.; Eckhardt, S.G. Assessment of the biological and pharmacological effects of the alpha3beta3 and alpha5beta5 integrin receptor antagonist, cilengitide (EMD 121974), in patients with advanced solid tumors. *Ann. Oncol.*, **2007**, *18*, 1400-07.
- [50] Chen, X.; Plasencia, C.; Hou, Y.; Neamati, N. Synthesis and biological evaluation of dimeric RGD peptide-Paclitaxel conjugate as

- a model for integrin-targeted drug delivery. *J. Med. Chem.*, **2005**, *48*, 1098-06.
- [51] Temming, K.; Raymond, M.S.; Molema, G.; Kok, R.J. RGD-based strategies for selective delivery of therapeutics and imaging agents to the tumour vasculature. *Drug Resist. Updat.*, **2008**, *8*, 381-402.
- [52] Beer, A.J.; Haubner, R.; Sarbia, M.; Goebel, M.; Luderschmidt, S.; Grosu, A.L.; Schnell, O.; Niemeyer, M.; Kessler, H.; Wester, H.J.; Weber, W.A.; Schwaiger, M. *Clin. Cancer Res.*, **2006**, *12*, 3942-49.
- [53] Haubner, R.; Weber, W.A.; Beer, A.J.; Vabuliene, E.; Reim, D.; Sarbia, M.; Becker, K.F.; Goebel, M.; Hein, R.; Wester, H.J.; Kessler, H.; Schwaiger, M. Non invasive visualization of the activated alphavbeta3 integrin in cancer patients by positron emission tomography and [18F]Galacto-RGD. *PLoS Med.*, **2005**, *2*, 70-73.
- [54] Livant, D.L.; Brabec, R.K.; Kurachi, K.; Allen, D.L.; Wu, Y.; Haaseth, R.; Andrews, P.; Ethier, S.P.; Markwart, S. The PHSRN sequence induces extracellular matrix invasion and accelerates wound healing in obese diabetic mice. *J. Clin. Invest.*, **2000**, *105*, 1537-45.
- [55] Khalili, P.; Arakelian, A.; Chen, G.; Plunkett, M.L.; Beck, I.; Parry, G.C.; DoOate, F.; Shaw, D.E.; Mazar, A.P.; Rabbani, S.A. A non-RGD-based integrin binding peptide (ATN-161) blocks breast cancer growth and metastasis *in vivo*. *Mol. Cancer Ther.*, **2006**, *5*, 2271-80.
- [56] Stoeltzing, O.; Liu, W.; Reinmuth, N.; Fan, F.; Parry, G.C.; Parikh, A.A.; McCarty, M.F.; Bucana, C.D.; Mazar, A.P.; Ellis, L.M. Inhibition of integrin alpha5beta1 function with a small peptide (ATN-161) plus continuous 5-FU infusion reduces colorectal liver metastases and improves survival in mice. *Int. J. Cancer*, **2003**, *104*, 496-503.
- [57] Funahashi, Y.; Sugi, N.H.; Semba, T.; Yamamoto, Y.; Hamaoka, S.; Tsukahara-Tamai, N.; Ozawa, Y.; Tsuruoka, A.; Nara, K.; Takahashi, K.; Okabe, T.; Kamata, J.; Owa, T.; Ueda, N.; Haneda, T.; Yonaga, M.; Yoshimatsu, K.; Wakabayashi, T. Sulfonamide derivative, E7820, is a unique angiogenesis inhibitor suppressing an expression of integrin alpha2 subunit on endothelium. *Cancer Res.*, **2002**, *62*, 6116-23.
- [58] Semba, T.; Funahashi, Y.; Ono, N.; Yamamoto, Y.; Sugi, N.H.; Asada, M.; Yoshimatsu, K.; Wakabayashi, T. An Angiogenesis Inhibitor E7820 Shows Broad-Spectrum Tumor Growth Inhibition in a Xenograft Model: Possible Value of Integrin alpha2 on Platelets as a Biological Marker. *Clin. Cancer Res.*, **2004**, *10*, 1430-38.
- [59] Haubner, R.; Schmitt, W.; Holzemann, G.; Goodman, S. L.; Jonczyk, A.; Kessler, H. Cyclic RGD peptides containing  $\beta$ -turn mimetics. *J. Am. Chem. Soc.*, **1996**, *118*, 7881-91.
- [60] Lohof, E.; Planker, E.; Mang, C.; Burkhart, F.; Dechantsreiter, M.A.; Haubner, R.; Wester, H.J.; Schwaiger, M.; Hölzemann, G.; Goodman, S.L.; Kessler, H. Carbohydrate Derivatives for Use in Drug Design: Cyclic alpha(v)-Selective RGD Peptides. *Angew. Chem. Int. Ed.*, **2000**, *39*, 2761-64.
- [61] Belvisi, L.; Colombo, L.; Manzoni, L.; Potenza, D.; Scolastico, C. Design, synthesis, conformational analysis and application of azabicycloalkane amino acids as constrained dipeptide mimics. *Synlett*, **2004**, *9*, 1449-71.
- [62] Colombo, L.; Di Giacomo, M.; Brusotti, G.; Sardone, N.; Angiolini, M.; Belvisi, L.; Maffioli, S.; Manzoni, L.; Scolastico, C. Stereoselective synthesis of 6,5-bicyclic reverse-turn peptidomimetics. *Tetrahedron*, **1998**, *54*, 5325-36.
- [63] Belvisi, L.; Bernardi, A.; Manzoni, L.; Potenza, D.; Scolastico, C. Conformational Analysis of Azabicycloalkane Amino Acid Scaffolds as Reverse-Turn Inducer Dipeptide Mimics. *Eur. J. Org. Chem.*, **2000**, 2563-69.
- [64] Integrin targeted Cyclopeptide ligands, their preparation and use, International Patent WO 2008/102233 A1.
- [65] Manzoni, L.; Bassanini, M.; Belvisi, L.; Motto, I.; Scolastico, C.; Castorina, M.; Pisano, C. Nonpeptide Integrin Antagonists: RGD Mimetics Incorporating Substituted Azabicycloalkanes as Amino Acid Replacements. *Eur. J. Org. Chem.*, **2007**, 1309-17.
- [66] Gibson, C.; Sulyok, G.A.G.; Hahn, D.; Goodman, S.L.; Holzemann, G.; Kessler, H. Nonpeptidic  $\alpha\beta$ 3 integrin antagonist libraries: on-bead screening and mass spectrometric identification without tagging. *Angew. Chem. Int. Ed.*, **2001**, *40*, 165-169.
- [67] Sulyok, G.A.G.; Gibson, C.; Goodman, S.L.; Holzemann, G.; Kessler, H. Solid-phase synthesis of a nonpeptide RGD mimetic library: new selective  $\alpha\beta$ 3 integrin antagonists. *J. Med. Chem.*, **2001**, *44*, 1938-50.
- [68] Goodman, S.L.; Holzemann, G.; Sulyok, G.A.G.; Kessler, H. Nanomolar Small Molecule Inhibitors for  $\alpha\beta$ 6,  $\alpha\beta$ 5, and  $\alpha\beta$ 3 Integrins. *J. Med. Chem.*, **2002**, *45*, 1045-51.
- [69] Agrez, M.; Chen, A.; Cone, R. I.; Pytela, R.; Sheppard, D. The alphav beta6 integrin promotes proliferation of colon carcinoma cells through a unique region of the beta 6 cytoplasmic domain. *J. Cell. Biol.*, **1994**, *127*, 547-556.
- [70] Jones, J.; Watt, F. M.; Speight, P. M. Changes in the expression of  $\alpha$ v integrins in oral squamous cell carcinomas. *J. Oral. Pathol. Med.*, **1997**, *26*, 63-68.
- [71] Kim, S.; Bell, K.; Mousa, S.A.; Varner, A. Regulation of angiogenesis *in vivo* by ligation of integrin  $\alpha$ 5 $\beta$ 1 with the central cell-binding domain of fibronectin. *Am. J. Pathol.* **2000**, *156*, 1345-62.
- [72] Benfatti, F.; Cardillo, G.; Fabbioni, S.; Galzerano, P.; Gentilucci, L.; Juris, R.; Tolomelli, A.; Baiula, M.; Spart, A.; Spanpanato, S. Synthesis and biological evaluation of non-peptide  $\alpha\beta$ 3/ $\alpha$ 5 $\beta$ 1 integrin dual antagonists containing 5,6-dihydropyridin-2-one scaffolds. *Bioorg. Med. Chem.*, **2007**, *15*, 7380-90.
- [73] Marinelli, L.; Meyer, A.; Heckmann, D.; Lavecchia, A.; Novellino, E.; Kessler, H. Ligand binding analysis for human  $\alpha$ 5 $\beta$ 1 integrin: strategies for designing new  $\alpha$ 5 $\beta$ 1 integrin antagonists. *J. Med. Chem.*, **2005**, *48*, 4204-07.
- [74] Heckmann, D.; Meyer, A.; Marinelli, L.; Zahn, G.; Stragies, R.; Kessler, H. Probing integrin selectivity: rational design of highly active and selective ligands for the  $\alpha$ 5 $\beta$ 1 and  $\alpha$ v $\beta$ 3 integrin receptor. *Angew. Chem. Int. Ed.*, **2007**, *46*, 3571-74.
- [75] Curley, G.P.; Blum, H.; Humphries, M. Integrin antagonists. *J. Cell. Mol. Life Sci.*, **1999**, *56*, 427-441.
- [76] Heckmann, D.; Meyer, A.; Laufer, B.; Zahn, G.; Stragies, R.; Kessler, H. Rational design of highly active and selective ligands for the  $\alpha$ 5 $\beta$ 1 integrin receptor. *ChemBioChem* **2008**, *9*, 1397-1407.
- [77] Stragies, R.; Osterkamp, F.; Zischinsky, G.; Vossmeier, D.; Kalhoff, H.; Reimer, U.; Zahn, G. Design and synthesis of a new class of selective integrin  $\alpha$ 5 $\beta$ 1 antagonists. *J. Med. Chem.*, **2007**, *50*, 3786-94.
- [78] Leonard, K.; Pan, W.; Anaclerio, B.; Gushue, J.M.; Guo, Z.; DesJarlais, R.L.; Chaikin, M.A.; Lattanze, J.; Cryslers, C.; Manthey, C.L.; Tomczuk, B.E.; Marugan, J.J. Non-peptidic  $\alpha$ , $\beta$  antagonists containing indol-1-yl propionic acids. *Bioorg. Med. Chem. Lett.*, **2005**, *15*, 2679-84.
- [79] Miller, W.H.; Alberts, D.P.; Bhatnagar, P.K.; Bondinell, W.E.; Callahan, J.F.; Calvo, R.R.; Cousins, R.D.; Erhard, K.F.; Heerding, D.A.; Keenan, R.M.; Kwon, C.; Manley, P.J.; Newlander, K.A.; Ross, S.T.; Samanen, J.M.; Uzinskas, I.N.; Venslavsky, J.W.; Yuan, C.C.-K.; Haltiwanger, R.C.; Gowen, M.; Hwang, S.; James, I.E.; Lark, M.W.; Rieman, D.J.; Stroup, G.B.; Azzarano, L.M.; Salyers, K.L.; Smith, B.R.; Ward, K.W.; Johanson, K.O.; Huffman, W.F. Discovery of Orally Active Nonpeptide Vitronectin Receptor Antagonists Based on a 2-Benzazepine Gly-Asp Mimetic. *J. Med. Chem.*, **2000**, *43*, 22-26.
- [80] <http://clinicaltrials.gov/ct2/show/study/NCT00689221?term=121974>.
- [81] <http://clinicaltrials.gov/ct2/show/NCT00352313?term=ATN161&rank=2>.
- [82] <http://clinicaltrials.gov/ct2/show/NCT00131651?term=ATN161&rank=1>.
- [83] <http://clinicaltrials.gov/ct2/show/NCT00078637?term=E7820&rank=2>.
- [84] <http://clinicaltrials.gov/ct2/show/NCT00309179?term=E7820&rank=1>.